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# 2 Phenetic and genetic structure of tsetse fly 3 populations (*Glossina palpalis palpalis*) in 4 southern Ivory Coast

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7 and Jean-Pierre Dujardin<sup>6\*</sup>

## 8 Abstract

9 **Background:** Sleeping sickness, transmitted by *G. p. palpalis*, is known to be present in the Ivory Coast. *G. p. palpalis*  
10 has recently been reported to occur in several places within the town of Abidjan, including: (i) the Banco forest, (ii) the  
11 Abobo Adjame University campus and (iii) the zoological park. Could these three places be treated sequentially, as  
12 separate tsetse populations, or should they be taken as one area comprising a single,  
13 panmictic population?

14 **Methods:** The amount of gene flow among these places provides strategic information for vector control. It was  
15 estimated by the use of both microsatellite DNA and morphometric markers. The idea was to assess the interest of the  
16 faster and much less expensive morphometric approach in providing relevant information about population  
17 structure. Thus, to detect possible lack of insect exchange between these neighbouring areas of Abidjan, we used  
18 both genetic (microsatellite DNA) and phenetic (geometric morphometrics) markers on the same specimens.  
19 Using these same markers, we also compared these samples with specimens from a more distant area of south Ivory  
20 Coast, the region of Aniassué (186 km north from Abidjan).

21 **Results:** Neither genetic nor phenetic markers detected significant differentiation between the three Abidjan *G. p.*  
22 *palpalis* samples. Thus, the null hypothesis of a single panmictic population within the city of Abidjan could not be  
23 rejected, suggesting the control strategy should not consider them separately. The markers were also in agreement  
24 when comparing *G. p. palpalis* from Abidjan with those of Aniassué, showing significant divergence between the two  
25 sites.

26 **Conclusions:** Both markers suggested that a successful control of tsetse in Abidjan would require the three Abidjan  
27 sites to be considered together, either by deploying control measures simultaneously in all three sites, or by a  
28 continuous progression of interventions following for instance the "rolling carpet" principle. To compare the  
29 geometry of wing venation of tsetse flies is a cheap and fast technique. Agreement with the microsatellite approach  
30 highlights its potential for rapid assessment of population structure.

## 31 Background

32 Tsetse flies (Diptera: Glossinidae) are the main vectors  
33 of trypanosomes (Kinetoplastida: Trypanosomatidae), which cause human and animal trypanosomiases  
34 in subsaharan Africa. These diseases have a considerable impact on public health and economic development

[1], although there are recent signs of a decline in incidence of the human disease following WHO-supported interventions based on case detection and treatment [2-4]. Vector control is an important complement to case detection and treatment, because reducing vector density can rapidly halt human trypanosomiasis transmission [5,6]. Vector control also remains the only strategy able to protect humans from acquiring a new infection [7].

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46 Tsetse populations may be reduced using a variety of  
47 techniques, including insecticide impregnated traps and  
48 targets, live-baits, sequential aerial spraying, and sterile  
49 male release [8-13]. However, in many cases when the  
50 control efforts have been stopped, the tsetse populations  
51 tend to recover due to flies surviving the initial interventions,  
52 or migrant flies coming from untreated regions, or  
53 both.

54 This has fueled debate as to whether in some instances  
55 “eradication” (defined by FAO as the creation of a tsetse  
56 free zone) may be more cost-effective than “suppression”  
57 where tsetse densities are reduced to a level minimizing  
58 the risk of disease transmission. Decisions on  
59 eradication or suppression strategies will be facilitated  
60 when the population structure within the target region,  
61 in particular the degree of genetic isolation of the target  
62 population from adjacent populations is clearly under-  
63 stood [14]. For isolated populations, eradication may be  
64 the most cost-effective strategy, as reported for *Glossina*  
65 *austeni* Newstead on Unguja Island, Zanzibar [9]. But  
66 for most mainland populations of tsetse, the geographical  
67 limits of target tsetse populations are less easily  
68 defined. Application of techniques that can detect pop-  
69 ulation isolation such as molecular or morphometric  
70 markers can guide decisions on the choice of control  
71 strategies [15-17]. Human and animal trypanosomiasis  
72 transmitted by *G. p. palpalis* are known to be present in  
73 Ivory Coast [4,18] and *G. p. palpalis* has been reported  
74 to occur within the city of Abidjan [19,20]. Due to  
75 its potential danger as a vector of human and animal  
76 trypanosomiasis, the Ivorian authorities now seek to  
77 control these tsetse flies in the affected area of Abid-  
78 jan, which includes the Banco forest, the University of  
79 Abobo Adjame and the zoological park. Tsetse have been  
80 found to be present in low to high densities in these 3  
81 sites, and were found infected by various trypanosome  
82 species [19].

83 To detect possible evidence of isolation between *G. p.*  
84 *palpalis* populations in the three affected areas within  
85 Abidjan, we used both genetic (microsatellite DNA) and  
86 phenetic (geometric morphometrics) markers on the  
87 same specimens, and compared these populations to *G. p.*  
88 *palpalis* populations from another area of southern Ivory  
89 Coast in the region of Aniassué. The idea was to assess  
90 the interest of the faster and much less expensive morpho-  
91 metric approach in providing relevant information about  
92 population structure.

93 The expected outcome of this study was to help the  
94 national control program to decide which is the best strat-  
95 egy of vector control in the town of Abidjan: can these  
96 three localities be treated sequentially (i.e. are the tsetse  
97 populations isolated between the three sites), or should  
98 they be taken as one area comprising a single, panmictic  
99 population?

## Results and discussion

### Microsatellite DNA markers

#### Within sample analyses

For the total sample ( $n = 141$ ) of genotyped tsetse, the seven microsatellite loci displayed 17 (Pgp1), 17 (PgP13), 14 (PgP24), 25 (B104), 19 (B110), 7 (C102), and 9 (GPCAG) alleles, respectively. The mean number of alleles was 9.71 (Banco), 11 (University) and 10.85 (Zoo) in Abidjan, and 10.00 in Aniassué. Mean observed heterozygosities were 0.68, 0.76 and 0.77 for Banco, University and Zoo, respectively, and 0.70 in Aniassué (no significant difference).

Overall  $F_{is}$  values were 0.12, 0.09 and 0.05 for Banco, University, and Zoo, significant at  $p < 0.0001$ ,  $p < 0.001$ , and  $p < 0.05$  respectively. In Aniassué,  $F_{is}$  was 0.15,  $p < 0.0001$ . The heterozygote deficit was mainly due to two loci (PgP1 and B110) for the three populations of Abidjan (Figure 1).

This suggested locus had specific technical problems (e.g. null alleles or short allele dominance), because when these loci were removed from the analysis,  $F_{is}$  values dropped to non-significant values (0.04, 0.00 and 0.03, respectively). Hence the null hypothesis of panmixia in Abidjan could not be rejected. In Aniassué,  $F_{is}$  on these 5 loci was 0.18 ( $p < 0.0001$ ), indicating consistent heterozygote deficiency. The heterozygote deficiency found in Aniassué confirmed earlier observations on *G. p. palpalis* in the forested areas of Ivory Coast, which attributed such deficiency to a combination of null alleles and genetic structuring at local scale due to Wahlund effects [21].

#### Genetic differentiation between samples

The mean  $F_{st}$  value for the 5 loci among the four populations was estimated at  $\theta = 0.017$  (CI95:  $0.011 < \theta < 0.023$ ),  $p < 0.0001$ . For the Abidjan samples it was  $\theta = 0.007$  (CI95:  $0.00150 < \theta < 0.01184$ ) and was not significant, meaning that most of the differentiation was due to differences between Aniassué and Abidjan. Looking at paired  $F_{st}$  values between sites (Table 1) confirmed this, since the highest (and significant) values always included Aniassué. Within Abidjan, there was a slight but non-significant trend for the population of Banco to diverge ( $F_{st} = 0.01$ ,  $p < 0.05$ ) from those of University and Zoo, whereas the latter two were genetically similar.

## Geometric morphometrics

### Size: centroid size

The specimens from Aniassué were significantly smaller compared to those from Abidjan, whereas within Abidjan there was no significant size difference between flies from the three sites (Figure 2).

### Shape variation

The first two discriminant factors derived from the shape variables showed that the polygon representing the

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136 T1

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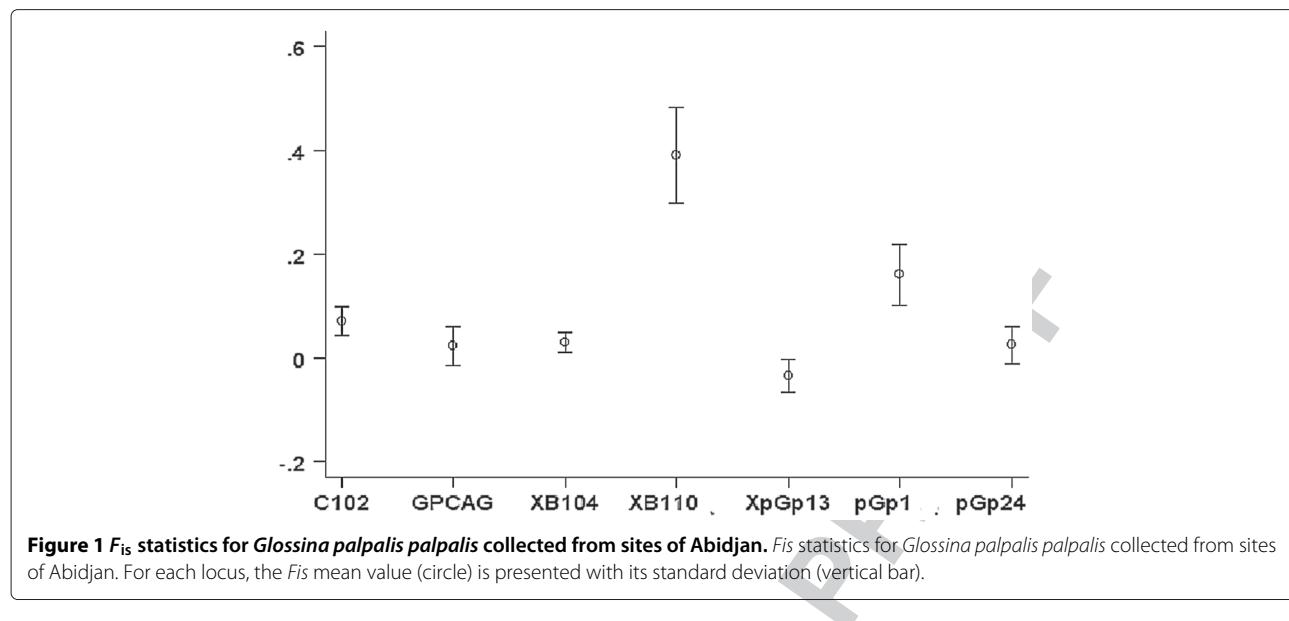
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147 F2

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Aniassué population tended to separate from the Abidjan sites (Figure 3). The reclassification tree, based on all three of the discriminant factors, clearly separated the Aniassué sample from those from Abidjan (Figure 4).

The Mahalanobis distances between the Abidjan samples (Table 1) were not significantly different, indicating an absence of shape differentiation, while the Mahalanobis distances from Aniassué were significantly larger ( $p < 0.007$ ) (Table 1).

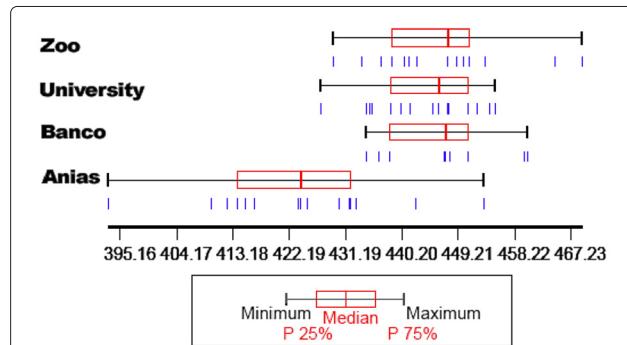
The validated reclassification scores confirmed this pattern, since Aniassué had the highest score (86%). However, in spite of the lack of significant differentiation within Abidjan, the reclassification score obtained for Banco (77%) was much higher than for the University (37%) and Zoo (33%), suggesting a relatively higher level of shape divergence in the Banco forest.

Correlation between metric and genetic distances was high. Regression of the Mahalanobis distances on the genetic distances indicated that 79% of the morphometric variation could be explained by the genetic variation (Figure 5).

#### Genetic and morphometric differentiation

From an epidemiological point of view, our study aimed at knowing whether tsetse populations from three sites in Abidjan could be considered to be isolated from each other. Such information is relevant for designing an adequate tsetse control strategy. For example, an insecticide application could be sequential in case of separation between sites, working on each site separately without risk of reinvasion to the next, or it should simultaneously cover all three sites if no evidence for separation is found.

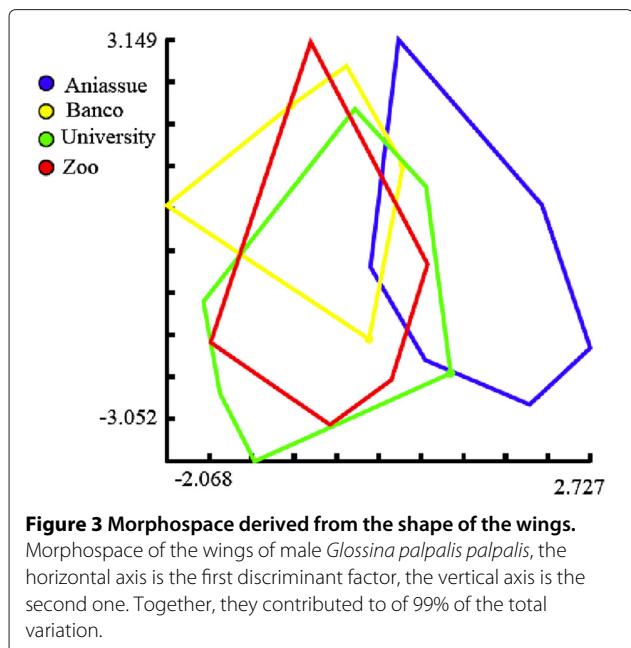
We used a population genetics approach [7] to analyse possible separation between the three Abidjan populations, comparing genetic and phenetic markers. Thus, the study also tested the potential of geometric morphometrics as a possible surrogate for molecular markers.



**Figure 2** Size variation of the wings. Variation of the centroid size of the wing of male *Glossina palpalis palpalis* according to localities. Anias, Aniassué. Each box shows the group median separating the 25th and 75th quartiles. Vertical bars under the boxes represent the wings. Units are pixels. P, percentile.

**Table 1** Metric and genetic distances between sites

Population 1	Population 2	Mahalanobis	$F_{ST}$
Aniassué	Banco	2.38	0.0221
Aniassué	University	1.93	0.0328
Aniassué	Zoo	1.98	0.0292
Banco	University	1.3	0.0121
Banco	Zoo	1.1	0.0113
University	Zoo	0.31	-0.0034
Pairwise metric (Mahalanobis) and genetic ( $F_{ST}$ ) distances between Banco, University, Zoo (Abidjan) and Aniassué.			

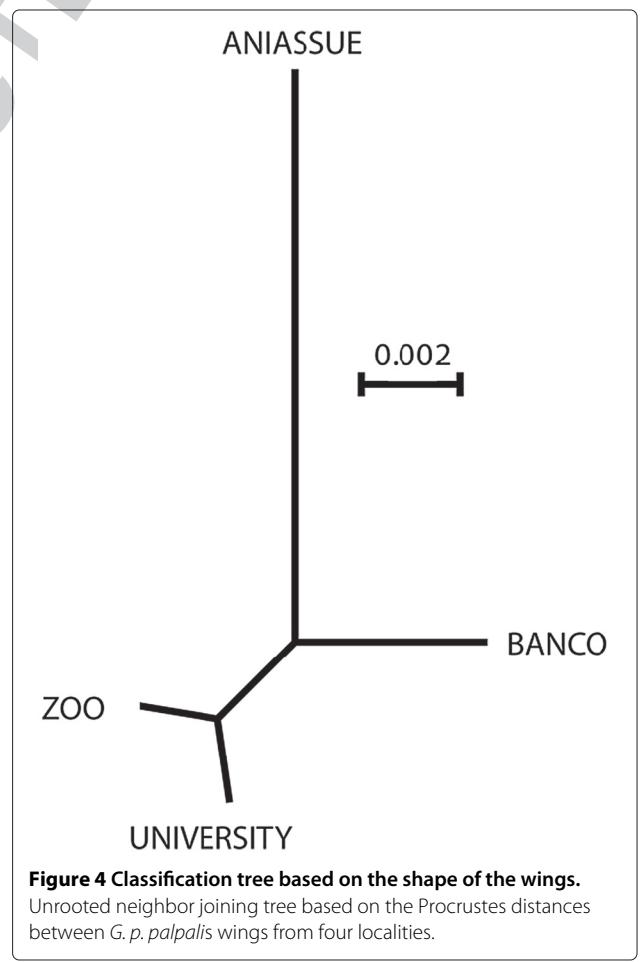


evidence for some morphometric specificity in the forest might be due to an environmental effect ("forest" versus "city"), although in tsetse most of the pre-imago development is relatively protected from external influences as tsetse larvae grow in the uterus of their mother during the three first stages, buffering morphometric variations against external influences [23].

Temperature and humidity do become influential factors at the time when pupae are in the soil. The effect has been studied for size (not shape), indicating that higher temperatures tend to result in smaller individuals [23], whereas increasing humidity tends to result in larger individuals [24]. It has been shown that the size of *G. p. palpalis* in forested areas of Ivory Coast is governed by seasonal climatic effects [25]. In Abidjan, no size difference was detected between sites, and given their proximity it seems likely that environmental factors acted uniformly on size.

#### Between Abidjan and Aniassué

By contrast, both molecular (microsatellite loci) and morphometric (centroid size and shape variables) data showed



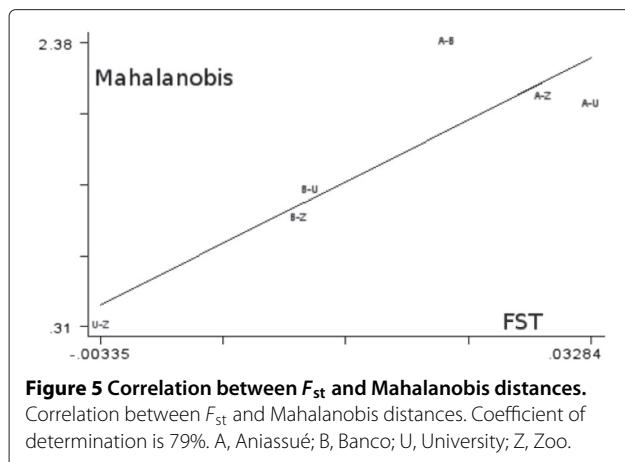
Both the phenetic (geometric morphometrics) and genetic (microsatellite loci) markers showed no evidence for differentiation between *G. p. palpalis* from sites within Abidjan, but both markers agreed in showing strong differentiation between individuals from Aniassué and those from Abidjan.

#### Within Abidjan

At the scale of Abidjan, our data showed that males from the three sites showed no genetic differentiation, and accordingly had similar metric properties (size and shape).

The microsatellite markers did not show any significant departure from the null hypothesis of panmixia, i.e. we did not observe any genetic differentiation between the 3 populations within Abidjan. There was however a slight, non significant trend for the population of Banco to diverge from the two others. A possible explanation is then a slow, on-going process for this population of Banco to have less genetic exchanges with the two others, due to urbanization which restricts tsetse movements. It may be possible, as observed in other studies in Burkina Faso, that the molecular markers used are not sensitive enough to detect it, since this is a recent, on-going phenomenon whereas what the molecular markers show is the result of a genetic history over several generations. This lack of sensitivity of molecular markers for recent genetic changes has already been observed in tsetse studies [22], and may be compensated by the use of morphometrics.

This idea is reflected by the much higher shape-based reclassification score obtained for Banco (77%), compared to the two other sites (37% and 33%). This indirect



gene flow at this scale. From a control perspective, this means that intervention against tsetse in any one site is likely to face reinvasion from the other two. This is different from a similar study conducted on the Loos archipelago, Guinea, which showed that tsetse populations (*G. palpalis gambiensis*) were isolated from the mainland and structured according to the island [15,34], which then allowed a sequential control strategy to be implemented [16,35]. Successful control of tsetse in Abidjan however, would require all three sites to be considered together (Figure 6), either by deploying control measures simultaneously in all three sites, or by a continuous progression of interventions - for example using barriers of impregnated traps and/or targets between sites (Figure 7) following the "rolling carpet" principle [36].

## Methods

### Study area

In Abidjan, the three study sites were the Banco forest (Banco), the University of Abobo Adjame (University) and the zoological park (Zoo). The Banco forest is in the north-western part of the city of Abidjan, at 5°N latitude and 4°W longitude. East of Banco are two small relicts of the forest which have now been substantially degraded by urbanisation: the Abobo Adjame University and the zoo of Abidjan. These three sites, although geographically close (less than 500 meters between sites), are separated by roads and urbanisation (Figure 8). For comparison, another study site was chosen near the town of Aniassué, about 186 km from Abidjan, in the Department of Abengourou, where *G. p. palpalis* occurs along the Comoé river. This region is characterized by forest degraded by wood

239 significant differences between tsetse from Abidjan and  
240 Aniassué. This was expected due to the geographic dis-  
241 tance between the two sites (186 km), to the differences of  
242 biotopes, and to the fact that the tsetse belt in South Ivory  
243 Coast is discontinuous as a consequence of anthropic  
244 pressure on habitats.

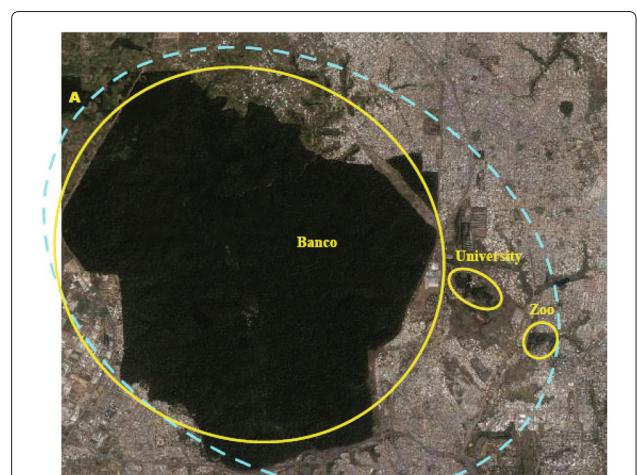
245 The tsetse from Aniassué were smaller than those from  
246 Abidjan. This was in agreement with both the slightly  
247 higher temperature [23] and dryer conditions [24,26] in  
248 Aniassué.

249 The differences between *G. p. palpalis* from Abidjan and  
250 Aniassué also involved shape, which may reflect genetic  
251 variations [23,27], especially when shape is allometry-free  
252 [28–30]. This was confirmed by differences found using  
253 microsatellite DNA markers. The parallel between phe-  
254 netic and genetic markers applied to natural populations is  
255 not uncommon [30]; for *G. p. gambiensis*, a similar parallel  
256 was observed in natural populations of different biotopes  
257 from West Africa [31], Guinea [15], Burkina Faso [32] and  
258 Senegal [17]. Here, 79% of the variance in Mahalanobis  
259 distance could be "explained" by genetic variation (com-  
260 pared to 50% in study by [17] study). This correlation does  
261 not imply a causal relationship, and could be attributed  
262 to both phenetic and genetic distances being related to  
263 geographical distances [33].

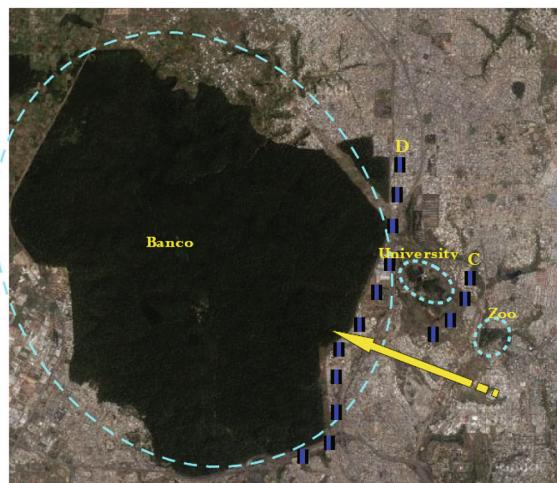
264 The heterozygote deficits found in Aniassué confirmed  
265 earlier observations on *G. p. palpalis* in the forested areas  
266 of Ivory Coast, which attributed such deficits to a combi-  
267 nation of null alleles and genetic structuring at local scale  
268 due to Wahlund effects [21].

## 269 Conclusions

270 How can the knowledge of population structure help  
271 to choose a control strategy? Since microsatellite and  
272 morphometric markers did not show significant differ-  
273 entiation between tsetse from the three sites in Abid-  
274 jan, there would appear to be no significant barrier to



**Figure 6 Eradication strategy.** Eradication strategy by controlling simultaneously the three sites. Blue dotted line: limits of the area to be treated simultaneously. Yellow curves: limits of target sites A: Relic forest of Anguededou not infested by tsetse flies.



**Figure 7** The “rolling carpet” principle. Eradication strategy in stages, site after site, but by creating barriers with traps or impregnated screens between Zoo and University (barrier C), University and Banco (barrier B), using the “rolling carpet” principle (Vreysen et al., 2007). The yellow arrow indicates the direction of the steps.

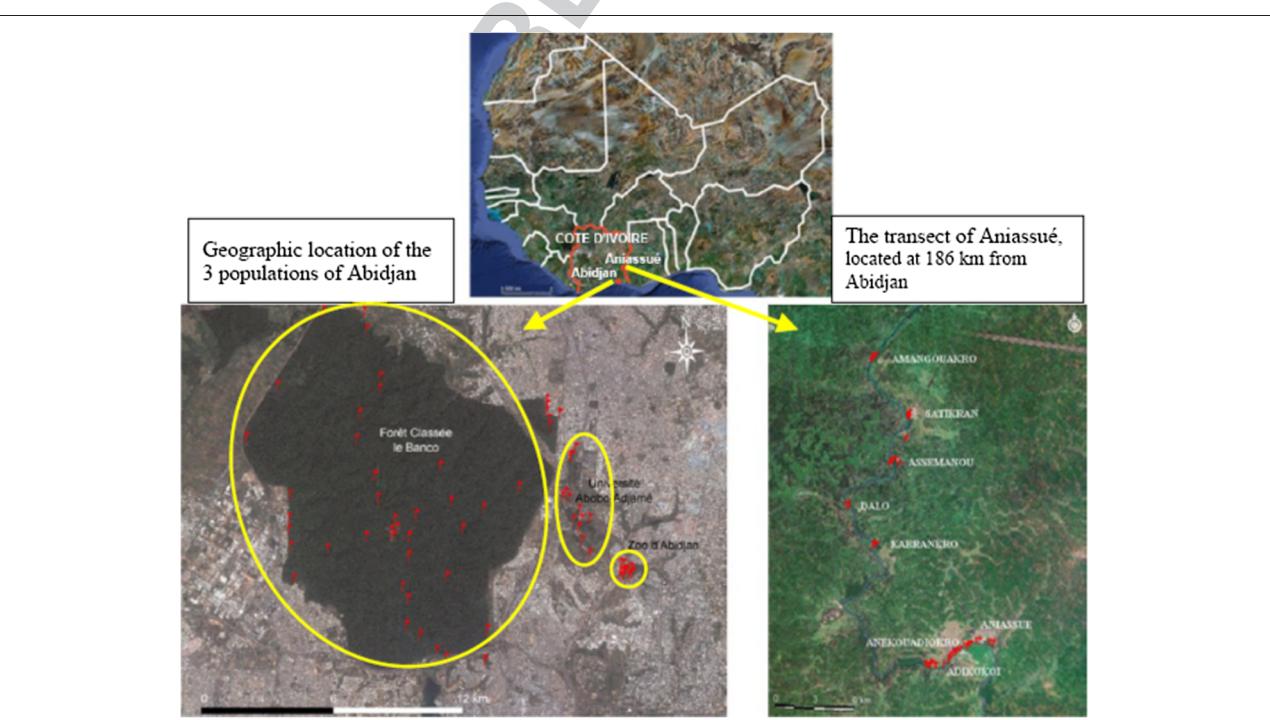
27.9°C), but relatively more variation in relative humidity (RH), which decreases from south (RH on average 90%) to north (RH between 60% and 70%). In both areas, there are two rainy and two dry seasons during a year [37].

#### Tsetse samples and microsatellite DNA markers

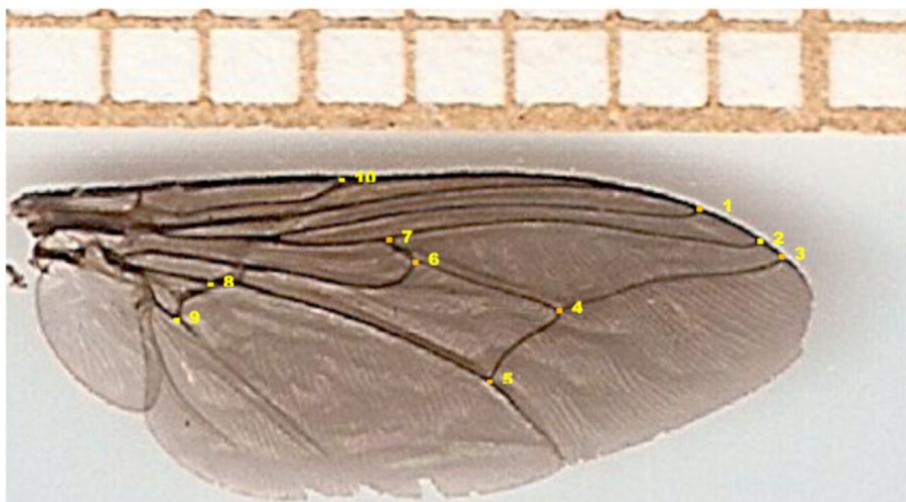
Tsetse flies were caught using Vavoua traps [38] in April 2007 in Aniassué and in October 2007 in Abidjan. In total from Abidjan 111 individual tsetse were analysed using microsatellite DNA markers: Banco (25 females (F), 11 males (M)); University (21 F, 17 M) and Zoo (21 F, 16 M). From Aniassué, 30 individuals were analysed (15 F and 15 M). Seven microsatellite markers were used (preceded by “X” for X-linked loci): Pgp1, XPgp13, Pgp24 [39], XB104, XB110, C102 (A. Robinson, FAO/IAEA, pers. com.) and GPCAG [40]. The samples were processed for Polymerase Chain Reaction (PCR) and genotyping on a 4300 DNA Analysis System from LI-COR (Lincoln, NE) as described in [34].

#### Population genetics analyses on molecular markers

Wright's F-statistics [41], the parameters most widely used to describe population genetic structure, were initially defined for a three-level hierarchical population structure (individuals, sub-populations, and total). In such a structure, three fixation indices or F-statistics can be defined.



**Figure 8** Geographic area of the study. Sampling sites of *Glossina palpalis palpalis* in Abidjan and Aniassué, Ivory Coast.



**Figure 9 Anatomical landmarks of the wing.** Ten landmarks at the junction of different veins in the wing of *Glossina palpalis palpalis*. Scale indicates millimeters.

337      $F_{is}$  is a measure of the inbreeding of individuals (hence I)  
338     resulting from non-random union of gametes within each  
339     sub-population (hence S).

340      $F_{st}$  quantifies the differentiation between subpopula-  
341     tions in the total population (hence S and T) as a  
342     measure of the relatedness between individuals result-  
343     ing from non-random distribution of individuals between  
344     sub-populations, relative to the total population.

345      $F_{it}$  is a measure of the inbreeding of individuals resulting  
346     both from non-random union of gametes within sub-  
347     populations, and from population structuring (deviation  
348     from panmixia of all individuals of the total population,  
349     hence I and T).

350     These F-statistics were estimated by Weir and Cocker-  
351     ham's unbiased estimators  $f$  (for  $F_{is}$ ),  $\theta$  (for  $F_{st}$ ) and  $F$  (for  
352      $F_{it}$ ) [42]. The significance of the F-statistics was tested by  
353     1000 random permutations in each case. The significance  
354     of  $F_{is}$  was tested by randomizing alleles between individ-  
355     uals within sub-samples. The significance of  $F_{st}$  was tested  
356     by randomizing individuals among sub-samples.

#### 357     Geometric morphometrics analyses

358     The tsetse specimens used for geometric morphometrics  
359     constituted a subsample of those on which the molec-  
360     ular analyses were done. Out of the 111 flies used for  
361     microsatellites, 55 had non-damaged wings allowing mor-  
362     phometric analyses. The analyses were conducted only on  
363     males, and focused on the right wing, which was generally  
364     the wing in best conditions. A total of 55 right wings of  
365     *G. p. palpalis* males (M) were used, i.e. 9 from Banco, 16  
366     from University, 15 from Zoo and 15 from Aniassué.

367     Wings were dry-mounted between two microscope  
368     slides and scanned at 1800 ppp at dimensions of 0.90

x 0.50 cm, using a multifunction scanner HP Deskjet F 369  
370 2180. From this picture, the coordinates of 10 landmarks 371  
372 (LM) defined by vein intersections were recorded for each 372 **F9**  
373 wing, by the same person in the same order (Figure 9). 373  
374 Repeatability was estimated at better than 80% (discussed 374  
375 elsewhere:[43]). 375

Raw coordinates were superimposed using the Generalized 375  
376 Procrustes Analysis (GPA) [44,45], producing one 376  
variable for size and 16 variables for shape. 377

The size variable was the isometric estimator known 378  
379 as centroid size (CS) derived from coordinate data 380  
381 and defined as the square root of the sum of the 382 squared distances between the center of the configura- 383  
384 tion of landmarks, and each individual landmark [46]. 385  
Statistical significance for size comparisons was esti- 385  
386 mated by 1,000 permutation tests [47] with Bonferroni 386  
correction. 387

The 16 shape variables were the "partial warps" (PW). 386  
To circumvent the problem of small sample sizes rela- 387  
tive to the large number of shape variables (16 PW), we 388  
used the first 6 principal components of the PW (relative 389  
warps, RW) as input for discriminant analyses, as these 390

**Table 2 Reclassification of tsetse individuals based on the shape of the wings**

Populations	Correctly assigned individuals	t2.1
Aniassué	13 / 15	86%
Banco	7 / 9	77%
University	6 / 16	37%
Zoo	5 / 15	37%
Validated reclassification of tsetse individuals based on the shape of their wings.		t2.2
		t2.3
		t2.4
		t2.5
		t2.6
		t2.7
		t2.8

represented 84% of the total variation and had the highest discriminatory power [48].

Mahalanobis distances [49] computed from these 6 RW were used to quantify shape divergence between groups (Figure 4) and the statistical significance was estimated by 1000 permutation tests [50] with Bonferroni correction.

Mahalanobis distances based re-classification scores were computed according to a validation procedure whereby each individual was assigned to its closest group without using that individual to help determine a group centre [33], although the computed shape variables did include that individual [43] (Table 2).

## Software

Collections of anatomical landmarks of the wings, general Procrustes analysis (GPA), multivariate and discriminant analyses, were performed using the CLIC package [43], freely available at <http://www.mpl.ird.fr/morphometrics/clic/index.html>. PHYLP software with "neighbor" module [51] and NJPLOT [52] were used to build the classification tree. The F-statistics from molecular data were estimated with Genetix [53] and Fstat 2.9.3.2 (updated from [54]). The overall G-test was used to estimate the significance of  $F_{st}$  with Fstat [55].

## Competing interests

The authors declare that they have no competing interests.

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## Authors contributions

Genetic techniques: SR, KA, LG, EKN'G, PS. Morphometric techniques: DK, HB-V, J-PD. Data analyses: DK, SR, PS, J-PD. Field collections: DK, MK, GA-Y. Text: DK, PS, J-PD, CJS. All authors read and approved the final version of the manuscript.

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## References

1. Louis FJ: **Les raisons techniques de la réémergence de la maladie du sommeil.** Médecine Tropicale 2001, **61**:425–431.
2. Schofield CJ, Kabayo JP: **Trypanosomiasis vector control in Africa and Latin America.** Parasit Vectors 2008, **1**:24 doi:10.1186/1756-3305-1-24.
3. Simarro PP, Jannin J, Cattand P: **Eliminating Human African Trypanosomiasis. Where do we stand and what comes next?** PLOS Med 2008, **5**:174–180. e55.
4. Simarro PP, Cecchi G, Paone M, Franco JR, Diarra A, Ruiz JA, Fèvre EM, Courten F, Mattioli RC, Jannin JG: **The Atlas of human African trypanosomiasis: a contribution to global mapping of neglected tropical diseases.** Int J Health Geographics 2010, **9**(57):pp18.
5. Rogers DJ: **Tsetse population dynamics and distribution: a new analytical approach.** J Anim Ecol 1979, **48**:825–849.
6. Laveissière C, Penchenier L: **Manuel de lutte contre la maladie du sommeil.** IRD Editions, Collection Didactiques 2005, **4**:pp273.
7. Solano P, Ravel S, De Meeùs T: **How can tsetse population genetics contribute to African trypanosomiasis control?** Trends Parasitol 2010, **26**(5):255–263.
8. Challier A, Eyraud M, Lafaye A, Laveissière C: **Amélioration du rendement du piège biconique pour glossines (Diptera: Glossinidae) par l'emploi d'un cône inférieur bleu.** Cah ORSTOM sér Ent Méd Parasitol 1977, **15**:283–286.
9. Vreysen MJB, Saleh KM, Ali MY, Abdulla AM, Zhu ZR, Juma KG, Dyck VA, Msangi AR, Mkonyi PA, Feldmann FU: **Glossina austeni (Diptera: Glossinidae) eradicated on the island of Unguja, Zanzibar, using the Sterile Insect Technique.** J Econ Entomol 2000, **93**:123–135.
10. Cuisance D, Itard J, Solano P, Desquesnes M, Frézil JL, Authié E: **Trypanosomoses. Méthodes de lutte.** In Editions Médicales Internationales Tec & Doc. Principales maladies infectieuses et parasitaires du bétail. Europe et Régions chaudes, volume 1. Paris: Lavoisier; 2003:139–165.
11. Kgori PM, Modo S, Torr SJ: **The use of aerial spraying to eliminate tsetse from the Okavango Delta of Botswana.** Acta Trop 2006, **99**:184–199.
12. Bouyer J, Solano P, Cuisance D, Itard J, Frézil JL, Authié E: **Control methods in Trypanosomiasis.** In Infectious and Parasitic Diseases of Livestock. Edited by Lefèvre P, BRUG C, Chermette J: Lavoisier Tec & Doc; 2010:Chap. 127.
13. Torr SJ, Solano P: **Olfaction in Glossina-host interactions: a tale of two tsetse.** In Olfaction in vector hosts interactions: Ecology and control of vector borne diseases, Volume 2. Edited by Takken BKW. Wageningen University, Netherlands, pp437; 2010:265–289.
14. Patterson JS, Schofield CJ: **Preliminary study of wing morphometry in relation to tsetse population genetics.** S Afr J Sci 2005, **101**:132–134.
15. Camara M, Caro-Riaño H, Ravel S, Dujardin JP, Hervouet JP, de Meeus T, Kagbadouno MS, Bouyer J, Solano P: **Genetic and morphometric evidence for isolation of a tsetse (Diptera: Glossinidae) population (Loos islands, Guinée).** J Med Entomol 2006, **43**(5):853–860.
16. Kagbadouno M, Camara M, Bouyer J, Hervouet JP, Morifaso O, Kaba D, Jamonneau V, Solano P: **Tsetse elimination: its interest and feasibility in the historical sleeping sickness focus of Loos islands, Guinea.** Parasite 2009, **16**:29–36.
17. Solano P, Kaba D, Ravel S, Dyer N, Sall B, Vreysen MJB, Seck MT, Darbyshire H, Gardes L, Donnelly MJ, De Meeùs T, Bouyer J: **Population genetics as a tool to select tsetse control strategies : suppression or eradication of Glossina palpalis gambiensis in the Niayes of Senegal.** PLoS NTD 2010, **4**(5):e692.
18. Kaba D, Djé NN, Courtin F, Oke E, Koffi M, Garcia A, Jamonneau V, Solano P: **L'impact de la guerre sur l'évolution de la THA dans le centre-ouest de la Côte d'Ivoire.** Trop Med Int Health 2006, **11**:136–143.
19. Allou K, Acapovi-Yao G, Kaba D, Bosson-Vanga H, Solano P, Ngoran KE: **Chorologie et infection par les trypanosomes de Glossina palpalis palpalis dans la forêt du Banco et ses reliques, Abidjan (Côte d'Ivoire).** Parasite 2009, **16**:289–295.
20. Keck N, Herder S, Kaba D, Solano P, Gomez J, Cuny G, Davoust B: **Epidemiology of canine trypanosomosis by cross-sectional study in a urban focus of Côte d'Ivoire.** Parasite 2009, **16**:305–308.
21. Ravel S, T DM, P DJ, Zézé DG, Gooding RH, Sané B, Dusfour I, G C, Solano P: **Different genetic groups occur within Glossina palpalis palpalis in the sleeping sickness focus of Bonon, Côte d'Ivoire.** Infection, Genet Evol 2007, **7**:116–125.
22. Solano P, Ravel S, De Meeùs T: **How can tsetse population genetics contribute to African Trypanosomosis control?** Trends Parasitol 2010, **26**:255–263 doi:10.1016/j.pt.2010.02.006.
23. Glasgow JP: **Selection for size in tsetse flies.** J Anim Ecol 1961, **30**:87–94.
24. Dejardin J, Maillot L: **Biométrie de la Glossine. Etude statistique des mensurations de l'aile dans diverses communautés (Glossina fuscipes quanzensis, Pires).** Revue Elev Méd. vét Pays Trop 1947, **17**:97–102.
25. Sané B, Solano P, Garcia A, Fournet F, Laveissière C: **Variation intraspécifique de la taille des ailes et du thorax chez Glossina palpalis palpalis en zone forestière de Côte d'Ivoire.** Rev Elev Vét Pays Trop 2000, **53**(3):245–248.

- 519 26. Rogers DJ, Randolph SE: **Mortality rate and population density of**  
520 **tsetse flies correlated with satellite imagery****Mortality rate and**  
521 **population density of tsetse flies correlated with satellite imagery.**  
522 *Nature* 1991, **351**:739–741.  
523 27. Falconer DS: **Introduction to quantitative genetics.** Longman, London,  
524 UK 1981, **2**:pp340.  
525 28. Dujardin JP, Le Pont F: **Geographic variation of metric properties**  
526 **within the Neotropical sandflies.** *Infect Gen Evol* 2004, **4**(4):353–359.  
527 29. Dujardin JP: **Morphometrics applied to Medical Entomology.** *Infection,*  
528 *Gen Evol* 2008, **8**:875–890.  
529 30. Dujardin JP: **Modern morphometrics of medically important insects.**  
530 In *Genetics and Evolution of Infectious diseases.* Edited by Tibayrenc M:  
531 Elsevier; 2011:pp749. ISBN: 978-0-12-384890-1, Chapter 16, 473–501.  
532 31. Solano P, De La Rocque S, Cuisance D, Geoffroy B, T DM, Cuny G, Duvillet  
533 G: **Intraspecific variability in natural populations of *Glossina palpalis***  
534 **gambiensis from West Africa, revealed by genetics and**  
535 **morphometrics analyses.** *Med Vet Entomol* 1999, **13**:401–407.  
536 32. Bouyer J, Ravel S, Dujardin JP, de Meeus T, Vial L, Thevenon S, Guerrini L,  
537 Sidibe I, Solano P: **Population structuring of *Glossina palpalis***  
538 **gambiensis (Diptera: Glossinidae) according to landscape**  
539 **fragmentation in the Mouhoun river, Burkina Faso.** *J Med Entomology*  
540 2007, **44**(5):788–795.  
541 33. Manly BFJ: *Multivariate Statistical Methods: A Primer.* London: Chapman &  
542 Hall; 1986. pp159. ISBN 0-412-28620-3.  
543 34. Solano P, Ravel S, Bouyer J, Camara M, Kabagdouno MS, Dyer N, Gardes L,  
544 Herault D, Donnelly MJ, De Meeûs T: **Population structures of insular**  
545 **and continental *Glossina palpalis gambiensis* in littoral Guinea.** *PLoS*  
546 *NTD* 2009, **3**(3):e392 doi:10.1371/journal.pntd.0000392.  
547 35. Kabagdouno M, Camara M, Bouyer J, Courtin F, F OM, J SC, Solano P:  
548 **Progress towards the eradication of tsetse from the Loos islands,**  
549 **Guinea.** *Parasit Vectors* 2011, **4**(1):18.  
550 36. Vreyen MJB, Robinson AS, Hendrichs J: *Area-wide Control of Insect Pests:*  
551 *From Research to Field Implementation.* Edited by IAEA. Dordrecht, The  
552 Netherlands: Springer; 2007.  
553 37. Anonyme: **Départements et districts de Côte d'Ivoire.** CECI, Groupe  
554 Inter-Commun 2005;pp420.  
555 38. Laveissière C, Grébaut P: **Recherches sur les pièges à glossines**  
556 **(Diptera: Glossinidae). Mise au point d'un modèle économique : le**  
557 **piège "Vavoua".** *Trop Med Parasitol* 1990, **41**:185–192.  
558 39. Luna C, Bonizzoni M, Cheng Q, Robinson AS, Aksoy L, Sand Zheng:  
559 **Microsatellite polymorphism in tsetse flies.** *J Med Entomol* 2001,  
560 **38**:376–381.  
561 40. Baker MD, Kafur ES: **Identification and properties of microsatellite**  
562 **markers in tsetse flies *Glossina morsitans sensu lato* (Diptera:**  
563 **Glossinidae).** *Mol Ecol Notes* 2001, **1**:234–236.  
564 41. Wright S: **The interpretation of population structure by F-statistics**  
565 **with special regard to system of mating.** *Evolution* 1965, **19**:395–420.  
566 42. Weir CC, B S, Cockerham: **Estimating F-statistics for the analysis of**  
567 **population structure.** *Evolution* 1984, **38**:1358–1370.  
568 43. Dujardin JP, Kaba D, Henry AB: **The exchangeability of shape.** *BMC Res*  
569 *Notes* 2010, **3**:266 doi:10.1186/1756-0500-3-266.  
570 44. Rohlf FJ: **Rotational fit (Procrustes) methods.** In *Proceedings of the,*  
571 *Michigan Morphometrics Workshop. Special Publication Number 2. The*  
572 *University of Michigan Museum of Zoology. Ann Arbor, MI,* pp380. Edited by  
573 Rohlf F, Bookstein F. University of Michigan Museums, Ann Arbor;  
574 1990:227–236.  
575 45. Rohlf FJ: **Morphometric spaces, shape components and the effects of**  
576 **linear transformations.** In *Advances in Morphometrics. Proceedings of the*  
577 *1993 NATO-ASI on Morphometrics.* Edited by Marcus LF, Corti M, Loy A,  
578 Naylor G, Slice D. New York: Plenum, Publ. NATO ASI, ser. A, Life Sciences;  
579 1996:117–129.  
580 46. Bookstein FL: *Morphometric Tools For Landmark Data. Geometry and*  
581 *Biology.* NY: Cambridge University Press; 1991.  
582 47. Caro-Riaño H, Jaramillo N, Dujardin JP: **Growth changes in *Rhodnius***  
583 **pallidus under simulated domestic and sylvatic conditions.**  
584 *Infection, Gen Evol* 2009, **9**(2):162–168.  
585 48. Baylac M, Frieß M: **Fourier descriptors, Procrustes superimposition,**  
586 **and data dimensionality: An exemple of cranial shape analysis in**  
587 **modern human populations.** Chicago: Kluwer; 2005:145–165. Chap. 6.  
588 49. Mahalanobis PC: **On the generalized distance in statistics.** *Proc Natl*  
589 *Inst Sci India* 1936, **2**:49–55.

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